

Potential artifacts and misinterpretations when evaluating the ecotoxicological effects of nanomaterials

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ABSTRACT

Engineered nanomaterials (ENMs) have significant commercial potential in a broad range of industries for consumer products as a result of their novel properties. However, these same properties may cause unexpected risks once ENMs are released into the environment either intentionally or unintentionally. Thus, standard methods are needed to accurately and reproducibly assess the potential risk of ENMs. One factor that limits the applicability of standard ecotoxicology test methods for use with ENMs is that the unique behaviors of ENMs may cause artifacts or misinterpretations in these tests as a result of their unique behaviors. We briefly discuss these artifacts and misinterpretations and provide an illustrative example.

Keywords: nanoparticle, nanoecotoxicology, standard test methods, artifacts, nanomaterials

1 INTRODUCTION

Nanotechnology promises exciting innovations in a broad range of fields, and nanomaterials have substantial potential for incorporation into consumer products. Nanomaterials are defined using the definition from the International Organization for Standardization (ISO): engineered nanomaterials (ENMs) are materials with any external dimension between 1 nm and 100 nm or having an internal surface structure in those dimensions [1, 2]; other agencies

such as the FDA may not necessarily operate under this strict definition. One issue that has limited the commercialization of ENM-containing products is their potential impacts on humans and the environment. Standard methods are needed for assessing the potential risks of ENMs, but the behaviors of ENMs differ substantially from those of traditional environmental pollutants such as hydrophobic organic chemicals and inorganic pollutants such as lead. Moreover, a literature review of the nanotechnology environmental health and safety literature showed that uncertainty in the applicability of current standard test methods for use with ENMs is the most frequently cited source of uncertainty [3].

One of the substantial differences between the behaviors of traditional environmental pollutants and ENMs during ecotoxicology testing is that ENMs may cause artifacts and misinterpretations during many of these tests. While there have been numerous review articles on the ecotoxicity of ENMs in organisms [4-18], the potential experimental artifacts and misinterpretations during these tests have received substantially less attention. For example, artifacts have been previously observed in nanoecotoxicology testing as a result of an unintended byproduct produced during the ENM dispersion process [19, 20] and from ENM interference with an assay reagent [21-27]. Misinterpretations in nanoecotoxicology testing are also possible if the effect observed is mistakenly attributed to nanoparticles when dissolved ions are actually the cause of the toxic effect. In a recently submitted review article [28],

we have systematically reviewed the potential artifacts and misinterpretations related to testing the potential ecotoxicological effects of ENMs. Potential artifacts were identified at every step of nanoeotoxicology testing: during the initial material synthesis or procurement and associated impurities [29-31], ENM storage [32-36], ENM dispersion [19, 20, 37, 38], unacknowledged indirect toxicity effects such as nutrient depletion [39-41], and during the toxicity assays [21-27, 42-44]. Test recommendations for how to avoid or minimize these artifacts and misinterpretations were also provided including a comprehensive list of potential control experiments and what they could test. In this proceedings paper, we will not reiterate the information from that review article, but rather briefly provide an example of an artifact observed as a result of coating desorption during storage which impacted the subsequent ecotoxicity measurement [36].

2 EXPERIMENTAL

The full experimental method is available in a previous publication [36]. Briefly, multiwall carbon nanotubes (MWCNTs) were treated with $\text{HNO}_3/\text{H}_2\text{SO}_4$ (v/v = 3:1), filtered, and rinsed with boiling water. These “3:1 MWCNTs” were then grafted with polyethyleneimine (PEI) as described previously [45]. MWCNTs were produced with positive (MWCNT-PEI), negative (MWCNT-PEI-Suc), or neutral (MWCNT-PEI-Ac) surface charges. These materials were thoroughly dialyzed and then stored at 4°C for several months.

Daphnia magna neonates (1 d to 2 d old) underwent immobilization tests at a range of concentrations (0 mg/L to 40 mg/L) for MWCNTs and for each of the three types of PEI-modified MWCNTs [46]. Five replicates of ten neonates in 20-mL vials were tested after 24 h and 48 h with 3:1 or PEI-coated MWCNTs spiked to artificial freshwater ($\text{CaCl}_2 \times 2\text{H}_2\text{O}$ 58.8 mg L⁻¹, $\text{MgSO}_4 \times 2\text{H}_2\text{O}$ 24.7 mg L⁻¹, NaHCO_3 13.0 mg L⁻¹, and KCl 1.2 mg L⁻¹; hardness $[\text{Ca}^{+2}] + [\text{Mg}^{+2}] = 0.5$ mM). Several additional experiments were conducted to explore the potential for artifacts to influence the immobilization results. Solutions of each modified MWCNT at the highest concentrations tested were filtered using ashless Whatman cellulose filters (2.5 µm, grade 42) and *Daphnia* were exposed to the filtrate. The PEI polymer by itself was also tested; PEI-Suc and PEI-Ac were not tested as a result of experimental challenges associated with the synthesis, purification, and identification of these polymers in the absence of their covalent bonding to MWCNTs prior to dialysis. In addition, all three types of MWCNT-PEIs were dialyzed, and their toxicity tested immediately after dialysis. The percentages of *Daphnia* immobilized after 24 h and 48 h of exposure were plotted against test concentrations and the data analyzed by statistical probit method (BioStat 2009, AnalystSoft) to calculate EC_{50} values (i.e., the

concentration at which 50 % of the *Daphnia* become immobilized) and their 95 % confidence limits.

3 RESULTS AND DISCUSSION

One important unexpected finding of this study was that, while the filtrate from the MWCNT-PEI-Ac and MWCNT-PEI-Suc did not cause immobilization, the filtrate from the MWCNT-PEIs caused 18% immobilization. This suggested that the PEI itself may exert a significant toxic effect on the neonates. When the PEI by itself was tested, the 24 h EC_{50} value was 19.3 mg/L which is within the range tested for the MWCNTs. When the MWCNTs were dialyzed immediately before the immobilization experiment, the toxicity for the MWCNT-PEI were significantly reduced (see Figure 1) while those for the MWCNT-PEI-Suc and MWCNT-PEI-Ac were unchanged. These results suggest that desorption of the PEI occurred to the MWCNT-PEI during storage which caused an overestimation of the MWCNT-PEI toxic effects. This type of result prevents direct assignment of the toxicity to the nanomaterial. This example demonstrates the value of conducting a filtrate-only control experiment to investigate potential toxic effects from compounds released from the ENMs.

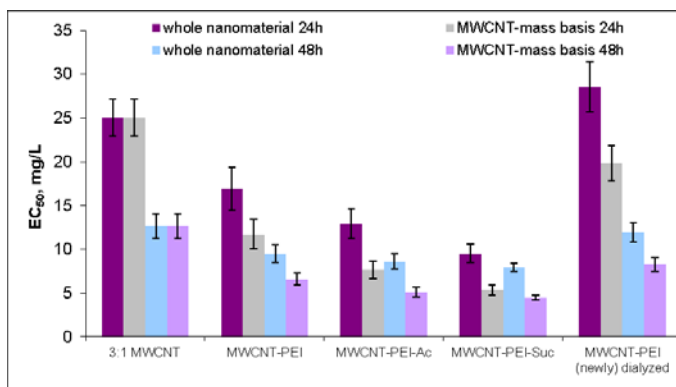


Figure 1: EC_{50} values for *Daphnia magna* exposed to regular and PEI modified MWCNTs. Five replicates of ten neonates were tested per concentration, and five to seven concentrations were tested for each type of MWCNT. Values are given after 24 and 48 h for the whole mass of the MWCNT with the PEI coating as indicated by “whole nanomaterial,” and on the basis of the MWCNT core by itself as indicated by “MWCNT-mass basis.” Values provided for the MWCNTs after they were recently dialyzed are marked “(newly) dialyzed.” Error bars represent the 95 % confidence intervals. This figure is modified and reprinted with permission from [36] copyright (2011) American Chemical Society.

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Certain commercial products or equipment is described in this paper in order to specify adequately the experimental

procedure. In no case does such identification imply recommendation or endorsement by the National Institute of Standards and Technology, nor does it imply that it is necessarily the best available for the purpose.

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REFERENCES

[1] ASTM (American Society for Testing Materials) International, E2456-06: standard terminology relating to nanotechnology. West Conshohocken, PA., 2006.

[2] ISO (International Organization for Standardization), TS 80004-1: nanotechnologies — vocabulary — Part 1: Core terms. Geneva, Switzerland, 2010.

[3] Grieger, K. D.; Hansen, S. F.; Baun, A., The known unknowns of nanomaterials: Describing and characterizing uncertainty within environmental, health and safety risks. *Nanotoxicol.* **2009**, *3*, (3), 1-U17.

[4] Klaine, S. J.; Alvarez, P. J. J.; Batley, G. E.; Fernandes, T. F.; Handy, R. D.; Lyon, D. Y., et al., Nanomaterials in the environment: Behavior, fate, bioavailability, and effects. *Environ. Toxicol. Chem.* **2008**, *27*, (9), 1825-1851.

[5] Handy, R. D.; Owen, R.; Valsami-Jones, E., The ecotoxicology of nanoparticles and nanomaterials: current status, knowledge gaps, challenges, and future needs. *Ecotoxicol.* **2008**, *17*, (5), 315-325.

[6] Kahru, A.; Dubourguier, H. C., From ecotoxicology to nanoecotoxicology. *Toxicology* **2010**, *269*, (2-3), 105-119.

[7] Handy, R. D.; Cornelis, G.; Fernandes, T.; Tsyusko, O.; Decho, A.; Sabo-Attwood, T., et al., Ecotoxicity test methods for engineered nanomaterials: Practical experiences and recommendations from the bench. *Environ. Toxicol. Chem.* **2012**, *31*, (1), 15-31.

[8] Holden, P. A.; Nisbet, R. M.; Lenihan, H. S.; Miller, R. J.; Cherr, G. N.; Schimel, J. P., et al., Ecological Nanotoxicology: Integrating Nanomaterial Hazard Considerations Across the Subcellular, Population, Community, and Ecosystems Levels. *Acc. Chem. Res.* **2013**, *46*, (3), 813-822.

[9] Baun, A.; Hartmann, N. B.; Grieger, K.; Kusk, K. O., Ecotoxicity of engineered nanoparticles to aquatic invertebrates: a brief review and recommendations for future toxicity testing. *Ecotoxicol.* **2008**, *17*, (5), 387-395.

[10] Peralta-Videa, J. R.; Zhao, L. J.; Lopez-Moreno, M. L.; de la Rosa, G.; Hong, J.; Gardea-Torresdey, J. L., Nanomaterials and the environment: A review for the

biennium 2008-2010. *J. Hazard. Mater.* **2011**, *186*, (1), 1-15.

[11] Tourinho, P. S.; van Gestel, C. A. M.; Loftis, S.; Svendsen, C.; Soares, A. M. V. M.; Loureiro, S., Metal-based nanoparticles in soil: Fate, behavior, and effects on soil invertebrates. *Environ. Toxicol. Chem.* **2012**, *31*, (8), 1679-1692.

[12] Kahru, A.; Ivask, A., Mapping the Dawn of Nanoecotoxicological Research. *Acc. Chem. Res.* **2012**, *46*, (3), 823-833.

[13] Pan, B.; Xing, B. S., Applications and implications of manufactured nanoparticles in soils: a review. *Eur. J. Soil Sci.* **2012**, *63*, (4), 437-456.

[14] Hou, W. C.; Westerhoff, P.; Posner, J. D., Biological accumulation of engineered nanomaterials: a review of current knowledge. *Environ. Sci. Proc. Imp.* **2013**, *15*, (1), 103-122.

[15] Ma, X. M.; Geiser-Lee, J.; Deng, Y.; Kolmakov, A., Interactions between engineered nanoparticles (ENPs) and plants: Phytotoxicity, uptake and accumulation. *Sci. Tot. Environ.* **2010**, *408*, (16), 3053-3061.

[16] Rico, C. M.; Majumdar, S.; Duarte-Gardea, M.; Peralta-Videa, J. R.; Gardea-Torresdey, J. L., Interaction of nanoparticles with edible plants and their possible implications in the food chain. *J. Agric. Food Chem.* **2011**, *59*, (8), 3485-3498.

[17] Miralles, P.; Church, T. L.; Harris, A. T., Toxicity, Uptake, and Translocation of Engineered Nanomaterials in Vascular plants. *Environ. Sci. Technol.* **2012**, *46*, (17), 9224-9239.

[18] Gogos, A.; Knauer, K.; Bucheli, T. D., Nanomaterials in Plant Protection and Fertilization: Current State, Foreseen Applications, and Research Priorities. *J. Agric. Food Chem.* **2012**, *60*, (39), 9781-9792.

[19] Oberdörster, E., Manufactured nanomaterials (Fullerenes, C₆₀) induce oxidative stress in the brain of juveniles largemouth bass. *Environ. Health Perspect.* **2004**, *112*, (10), 1058-1062.

[20] Henry, T. B.; Menn, F. M.; Fleming, J. T.; Wilgus, J.; Compton, R. N.; Saylor, G. S., Attributing effects of aqueous C₆₀ nano-aggregates to tetrahydrofuran decomposition products in larval zebrafish by assessment of gene expression. *Environ. Health Perspect.* **2007**, *115*, (7), 1059-1065.

[21] Worle-Knirsch, J. M.; Pulskamp, K.; Krug, H. F., Oops they did it again! Carbon nanotubes hoax scientists in viability assays. *Nano Lett.* **2006**, *6*, (6), 1261-1268.

[22] Holder, A. L.; Goth-Goldstein, R.; Lucas, D.; Koshland, C. P., Particle-Induced Artifacts in the MTT and LDH Viability Assays. *Chem. Res. Toxicol.* **2012**, *25*, (9), 1885-1892.

[23] Monteiro-Riviere, N. A.; Inman, A. O.; Zhang, L. W., Limitations and relative utility of screening assays to assess engineered nanoparticle toxicity in a human cell line. *Toxicol. Appl. Pharmacol.* **2009**, *234*, (2), 222-235.

- [24] Monteiro-Riviere, N. A.; Inman, A. O., Challenges for assessing carbon nanomaterial toxicity to the skin. *Carbon* **2006**, *44*, (6), 1070-1078.
- [25] Davis, R. R.; Lockwood, P. E.; Hobbs, D. T.; Messer, R. L. W.; Price, R. J.; Lewis, J. B., et al., In vitro biological effects of sodium titanate materials. *J. Biomed. Mat. Res. Part B Appl. Biomater.* **2007**, *83B*, (2), 505-511.
- [26] McCormack, T. J.; Clark, R. J.; Dang, M. K. M.; Ma, G. B.; Kelly, J. A.; Veinot, J. G. C., et al., Inhibition of enzyme activity by nanomaterials: Potential mechanisms and implications for nanotoxicity testing. *Nanotoxicol.* **2012**, *6*, (5), 514-525.
- [27] Xia, T.; Hamilton, R. F.; Bonner, J. C.; Crandall, E. D.; Elder, A.; Fazlollahi, F., et al., Interlaboratory Evaluation of in Vitro Cytotoxicity and Inflammatory Responses to Engineered Nanomaterials: The NIEHS Nano GO Consortium. *Environ. Health Perspect.* **2013**, *121*, (6), 683-690.
- [28] Petersen, E. J.; Henry, T. B.; Zhao, J.; MacCusprie, R. I.; Kirschling, T. L.; Dobrovolskaia, M. A., et al., Identification and avoidance of potential artifacts and misinterpretations in nanomaterial ecotoxicity measurements. **2014**, accepted.
- [29] Jakubek, L. M.; Marangoudakis, S.; Raingo, J.; Liu, X. Y.; Lipscombe, D.; Hurt, R. H., The inhibition of neuronal calcium ion channels by trace levels of yttrium released from carbon nanotubes. *Biomaterials* **2009**, *30*, (31), 6351-6357.
- [30] Liu, X. Y.; Gurel, V.; Morris, D.; Murray, D. W.; Zhitkovich, A.; Kane, A. B., et al., Bioavailability of nickel in single-wall carbon nanotubes. *Adv. Mat.* **2007**, *19*, (19), 2790-2796.
- [31] Hull, M. S.; Kennedy, A. J.; Steevens, J. A.; Bednar, A. J.; Weiss, C. A.; Vikesland, P. J., Release of metal impurities from carbon nanomaterials influences aquatic toxicity. *Environ. Sci. Technol.* **2009**, *43*, (11), 4169-4174.
- [32] Liu, J. Y.; Hurt, R. H., Ion Release Kinetics and Particle Persistence in Aqueous Nano-Silver Colloids. *Environ. Sci. Technol.* **2010**, *44*, (6), 2169-2175.
- [33] Liu, J. Y.; Sonshine, D. A.; Shervani, S.; Hurt, R. H., Controlled Release of Biologically Active Silver from Nanosilver Surfaces. *Acs Nano* **2010**, *4*, (11), 6903-6913.
- [34] Dobias, J.; Bernier-Latmani, R., Silver Release from Silver Nanoparticles in Natural Waters. *Environ. Sci. Technol.* **2013**, *47*, (9), 4140-4146.
- [35] Gorham, J. M.; B., R. A.; Lipka, K. A.; MacCusprie, R. I.; Hematti, A.; Holbrook, R. D., Storage Wars: How citrate capped silver nanoparticle suspensions are affected by not-so-trivial decisions. *J. Nano. Res.* **2014**, in press.
- [36] Petersen, E. J.; Pinto, R. A.; Mai, D. J.; Landrum, P. F.; Weber, W. J., Jr., Influence of polyethyleneimine graftings of multi-walled carbon nanotubes on their accumulation and elimination by and toxicity to *Daphnia magna*. *Environ. Sci. Technol.* **2011**, *45*, (3), 1133-1138.
- [37] Betts, J. N.; Johnson, M. G.; Rygiewicz, P. T.; King, G. A.; Andersen, C. P., Potential for metal contamination by direct sonication of nanoparticle suspensions. *Environ. Toxicol. Chem.* **2013**, *32*, (4), 889-893.
- [38] Wang, R.; Hughes, T.; Beck, S.; Vakil, S.; Li, S.; Pantano, P., et al., Generation of toxic degradation products by sonication of Pluronic(R) dispersants: implications for nanotoxicity testing. *Nanotoxicol.* **2013**, *7*, (7), 1272-1281.
- [39] Atha, D. H.; Wang, H. H.; Petersen, E. J.; Cleveland, D.; Holbrook, R. D.; Jaruga, P., et al., Copper Oxide Nanoparticle Mediated DNA Damage in Terrestrial Plant Models. *Environ. Sci. Technol.* **2012**, *46*, (3), 1819-1827.
- [40] Petersen, E. J.; Nelson, B. C., Mechanisms and measurements of nanomaterial-induced oxidative damage to DNA. *Anal. Bioanal. Chem.* **2010**, *398*, 613-650.
- [41] Nel, A.; Xia, T.; Madler, L.; Li, N., Toxic potential of materials at the nanolevel. *Science* **2006**, *311*, (5761), 622-627.
- [42] Horst, A. M.; Vukanti, R.; Priester, J. H.; Holden, P. A., An Assessment of Fluorescence- and Absorbance-Based Assays to Study Metal-Oxide Nanoparticle ROS Production and Effects on Bacterial Membranes. *Small* **2013**, *9*, (9-10), 1753-1764.
- [43] Weijie, W.; Yeow, J. T. W.; Van Dyke, M. I. In *Size-dependent PCR inhibitory effect induced by gold nanoparticles*, Engineering in Medicine and Biology Society, 2009. EMBC 2009. Annual International Conference of the IEEE, 3-6 Sept. 2009, 2009.
- [44] Hartmann, N. B.; Engelbrekt, C.; Zhang, J.; Ulstrup, J.; Kusk, K. O.; Baun, A., The challenges of testing metal and metal oxide nanoparticles in algal bioassays: titanium dioxide and gold nanoparticles as case studies. *Nanotoxicol.* **2013**, *7*, (6), 1082-1094.
- [45] Shen, M. W.; Wang, S. H.; Shi, X. Y.; Chen, X. S.; Huang, Q. G.; Petersen, E. J., et al., Polyethyleneimine-mediated functionalization of multiwalled carbon nanotubes: Synthesis, characterization, and in vitro toxicity assay. *J. Phys. Chem. C* **2009**, *113*, (8), 3150-3156.
- [46] Organization for Economic Cooperation and Development. 2004. *Daphnia* sp. Acute Immobilisation Test. OECD Guideline 202. Paris, F.